

Journal of Global Trends in Pharmaceutical Sciences



ISSN-2230-7346

REVIEW ON THE ANALYTICAL METHODS FOR THE ESTIMATION OF GARENOXACIN IN PHARMACEUTICAL DOSAGE FORMS

M. Ramakrishna, J. N. Suresh Kumar, B. Gowri bai, Ch. Sai Venkata Chandu, G. Vishnu, K. Nandini Devi, L. Jeevani

Narasaraopeta Institute of pharmaceutical sciences, Narasaraopet, Andhra Pradesh, 522601, India.

*Corresponding author E-mail: jeenilachi77@gmail.com

ARTICLE INFO

ABSTRACT

Key words:

Garenoxacin, analytical method development, method validation, method validation, reported methods, spectrometric methods.



This review explores Garenoxacin, a novel des-fluoro-6-quinolone antibiotic, highlighting its analytical methodologies. Garenoxacin exhibits broad spectrum activity against gram +ve & gram -ve organisms, including drug resistant strains. Its mechanism involves dual inhibition of DNA gyrase and topoisomerase IV, making it a potent candidate for the treating skin and soft tissue infections (SSSIs), respiratory tract infections, and other microbial conditions.. Analytical techniques such as HPLC (Concentration Range: 2–12 µg/mL, Wavelength: 280 nm, Stationary Phase: C18 (Princeton SPHERE ULTIMA, 250×4.6 mm, 5 µm), Mobile Phase: Acetonitrile: Water (60:40 v/v), ~0.1% orthophosphoric acid, Flow Rate: 1.0 mL/min, Detector: UV Diode Array (PDA)), RPHPLC, (chromatographic conditions used in stationary phase zorbax eclipse XDB C18 250×4.6mm, 5m, mobile phase 0.1% orthophosphoric acid and acetonitrile in the ration of 50:50 (%v\v) and flow rate was maintained at 1.0ml\min, detection wavelength 240nm concentration range of 0.04 to 4µg/ml),and UVspectrophotometry (Concentration Range: 3-18 µg/mL, Wavelength: 295 nm, Stationary Phase: Not applicable (UV method), Mobile Phase: Distilled water (aqueous solution) are discussed in the context of Garenoxacin quantification and validation, with emphasis on ICH and QbD guidelines. Clinical cases and studies underscore its effectiveness, safety and occasional adverse reactions, including hypersensitivity and fixed drug eruptions. The review also includes method development for pharmacokinetic studies and dosage form analysis, establishing garenoxacin's significance in therapeutic and analytical fields.

INTRODUCTION

Garenoxacin is considered as both API (bulk drug) and tablet formulation (solid oral dosage form). This study was carried out to assess the purity, quality and efficacy of using the new generation Garenoxacin these are the methods which effect the garenoxacin as: HPLC (High Performance Liquid Chromatography Effect of Garenoxacin: It has

polar functional groups (carboxylic acid, piperazinyl, quinolone core), so it interacts well with reversed-phase C18 columns. Retention time depends on pH (since garenoxacin has ionizable groups). Detection: UV detection at ~ 270–295 nm due to quinolone chromophore. Mobile Phase: Usually a mixture of buffer (phosphate, acetate) and organic solvent (acetonitrile/methanol). UHPLC (Ultra High

Performance Liquid Chromatography) Effect of Garenoxacin: Same principles as HPLC, but smaller particle size columns (≤2 µm) are used. Garenoxacin gives sharper peaks, shorter run time, higher sensitivity compared to HPLC. Advantage: More efficient separation and reduced solvent consumption. Spectroscopy (MS / LC-MS/MS) Effect of Garenoxacin: Because it contains nitrogen and fluorine atoms, garenoxacin ionizes well in Electrospray Ionization (ESI), usually in positive ion mode [M+H]+. Molecular ion peak: $m/z \sim 402 (M+H)+$ depending on the method. Useful for pharmacokinetic studies, metabolite identification, and trace level quantification. UV-Visible Spectrophotometer Effect of Garenoxacin: The quinolone nucleus has strong UV absorbance. Amax around 272-290 nm. Can be used for simple estimation in bulk and formulations (though less specific than HPLC/LC-MS). Limitation: Cannot distinguish garenoxacin degradation products or excipients as effectively **HPLC** as or LC-MS. HPLC/UHPLC → Used for assay, stability studies, dissolution. MS (LC–MS/MS) → highly sensitive, used for pharmacokinetics, analysis. plasma concentration spectrophotometer → Quick, simple method but less selective.

PHYSICAL AND CHEMICAL PROPERTY:

Table. No 1: Physical and chemical properties of Garenoxacin

of Garenoxaciii	
Chemical name	1-Cyclopropyl-8-
	(difluoromethoxy)-7-[(1R)-1-
	methyl-2,3dihydro-1H-
	isoindol-5-yl]-4-oxo-1,4-
	dihydroquinoline-3-
	carboxylic acid mono
	methane sulfonate
CAS Number	194804-75-6
Molecular	C23H20F2N2O4CH4O3S
formula	
Molecular	522.52g mol
weight	
Half life	20 hours
Solubility	Insoluble in water, sparingly
	soluble in methanol, very
	soluble in acetonitrile DMSO
Category	Antibiotic
BCS	Class II

CHEMICAL STRUCTURE OF GARENOXACIN:

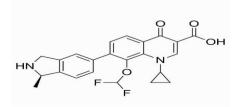


Fig.1: Chemical structure of garenoxacin

ANALYTICAL METHOD DEVELOPMENT:

The process of developing an accurate assay to ascertain the composition of formulations is known as analytical method development. Developing an analytical approach entails determining whether it is appropriate for use in laboratory. Analytical must adhere to GMP and GLP procedures and full fill ICH guide lines approval requirements^[18]

HIGH PERFORMANCE LIQUID CHROMATOGRAPHY (HPLC):

In Garenoxacin the HPLC method was introduced by A. P. Edlabadkar and A. P. Rajput in 2017 and 2018. HPLC method for optimization of development and validation of garenoxacin Mesylate in bulk and tablets.

Originally called high pressure liquid chromatography, HPLC is a technique used in analytical chemistry to separate, identify, and quantify specific components in mixtures among other things; the mixtures may originate from liquid solutions that have been dissolved for biological, environmental, medicinal, chemical or dietary sources. [19]

Optimised condition in HPLC

Flow rate - 1.0 ml \ min

Detection wavelength – 280nm

Stationary phase\ column - Princeton SPHERE ULTIMA C18, 280nm, 5µm particle size

Column- C18 column

Mobile phase- formic acid in water (0.1% v/v): methanol [(60:40) % v/v]

ULTRA-HIGH PERFORMANCE LIQUID CHROMATOGRAPHY (UHPLC):

Particles smaller than the 2.5-5um ones commonly employed in HPLC can be held in columns used in UHPLC. The fundamental premises of UHPLC, which operates under the same assumptions as HPLC that is efficiency and consequently, resolution accretion increase as column packing particle size decreases.

Smaller particle separations in columns result in increased efficiency per unit of time; however, efficiency cannot be reduced by higher mobile phase flow rates or linear velocities. Following characteristics, it is possible to achieve unprecedented levels of peak resolution and delicate particle speed. Linear velocity depends on the size of the particles packed into the analytical column. It is well known according to Van Deemter that the effectiveness of equations chromatographic process is directly correlated with the reduction in particle size. His modelcharacterized band broadening demonstrates that the relationship between the height equivalent of a theoretical plate (HETP) and linear velocity depends on analytical column.

REVERSE PHASE HIGH PERFORMANCE LIQUID CHROMATOGRAOHY (RPHPLC):

In Garenoxacin the RPHPLC was introduced by Ajitha Azhakesannn, Sujatha K, Abbulu K in April 2019. Bioanalytical method development and validation of garenoxacin Mesylate in human plasma by RP-HPLC.

A polar mobile phase and non-polar stationary phase are used in reversed phase liquid chromatography (RPLC) a kind of liquid chromatography, to separate organic molecules. The majority of separations and analyses carried out with HPLC in recent years have been carried out in the reversed phase mode. The components of the sample are kept in the system in reversed phase mode to a greater extent if they are hydrophobic. [21,22]

Stationary phase – zorbox eclipse XDB C18 (250×4.6mm, 5m)

Mobile phase -0.1% orthophosphoric acid and acetonitrile

Ratio -50.50 (%v\v)

Flow rate − 1.0 ml\min

Detection wavelength – 240nm

Retention time -4.0 min

Concentration range - 0.04-4 µg\ml

MASS PECTROSCOPY:

D. A. Gajjar and colleagues (Bristol-Myers Squibb) used LC-MS/MS (mass spectrometry) for garenoxacin quantification in humans in 2003.

Mass percentage of Carbon (C), Hydrogen (H), Fluorine (F), Nitrogen (N), and Oxygen (O) in

garenoxacin is 64.784%, 4.728%, 8.911%, and 15.008%. [23]

UV SPECTROPHOTOMETER:

Developed by: A. P. Edlabadkar & A. P. Rajput Methods introduced: Four UVspectrophotometric approaches (zero-order and first-order derivative, both with absorbance and AUC) for garenoxacin Mesylate assay

Publication source: Der Pharmacia Letters (mid-2010s; estimated ~2018)

Beer-lamberts range (ug/ml): 03-18 ug/ml

Spectral slit width: 1nm

Solvent: water

Lamda max: wave length maxima of garenoxacin with respected to adopted method like zero order, zero order-AUC, first order derivative, first order derivative AUC can be found as 274nm, 256.50-286nm, and 277.50-300nm^{· [24]}

REPORTED METHOD FOR GARENOXACIN:

In 2016, A. P. Edlabadkar, and A. P. Rajput Der Pharmacia Letter on "Development Simple **UV-Spectrophotometry** Four Estimation of Garenoxacin Methods for Mesylate in Bulk Material and Pharmaceutical Formulation." It details the development and validation of four UV-spectrophotometric methods for quantifying Garenoxacin Mesylate (GRN), a quinolone antibiotic used to treat respiratory and urinary tract infections, in bulk and tablet forms, using distilled water solutions were created using distilled water as a solvent. The study was published in 2021 by Aiitha Azhakesannn A and her colleagues. In order to develop and validate a novel stabilityindicating method based on QbD for the assay and dissolution of garenoxacin in tablets. It describes how a Quality by Design (QbD) methodology was used to create and validate a novel stability-indicating method for the assay and dissolving of garenoxacin in 200 mg garenoxacin tablets. The des-fluoro quinolone antibiotic garenoxacin Mesylate, which is sold under the brand name Geninax in Japan, has a broad-spectrum effect on antimicrobial resistance. With peak purity verified under stressful circumstances, the RP-HPLC test technique was linear, robust, accurate (99.8-100.5% recovery), and exact (RSD \leq 0.48%). Variations in API particle size had no effect on the dissolving method's consistent drug release ($Q \ge 80\%$ at 30min).

The study was published in 2021 by Ajitha Azhakesannn A and her colleagues. In order to develop and validate a novel stabilityindicating method based on QbD for the assay and dissolution of garenoxacin in tablets. It describes how a Quality by Design (QbD) methodology was used to create and validate a novel stability-indicating method for the assay and dissolving of garenoxacin in 200 mg garenoxacin tablets. The des-fluoro quinolone antibiotic garenoxacin Mesylate, which is sold under the brand name Geninax in has a broad-spectrum effect on antimicrobial resistance. With peak purity verified under stressful circumstances, the RP-HPLC test technique was linear, robust, accurate (99.8-100.5% recovery), and exact (RSD \leq 0.48%). Variations in API particle size had no effect on the dissolving method's consistent drug release ($Q \ge 80\%$ at 30min).

Spectrometric methods

Sakariya SV, mardia RB, Chauhan SP, development Sahagia BN of different spectrometric methods for the estimation of Mesylate garenoxacin in bulk and pharmaceutical formulations this was published by 2015, a different spectrometric method has developed and validate been for determination of GRA in tablet dosage form, these are the solvents are used in the preparation of garenoxacin: garenoxacin Mesylate 0.1N HCL, 0.1N NaOH, distilled water. These are the concentration of HCL and $10-50\mu g\mbox{ml}$. maximum are the wavelength was observed at 347nm.

Linearity: 10-50µg\ml

LOD: 0.32µg\ml LOQ: 1.09µg\ml

These are concentrations of HCL and distilled water are 10-50 μ g\ml, the maximum wavelength was observed at 348nm.

Linearity: 10-50µg\ml LOD: 0.46µg\ml LOQ: 1.55µg\ml CONCLUSION:

Garenoxacin analytical methods such RP-HPLC, UHPLC, UVand as spectrophotometry have been successfully developed and validated for its quantification, guidelines. **ICH** and QbD adhering to broad-spectrum Garenoxacin. a fluoroquinolone antibiotic, can be effectively analyzed by several modern analytical

techniques. UV spectrophotometry provides a simple and rapid method for its estimation, while HPLC and UHPLC offer accurate, reliable determination precise, and pharmaceutical formulations and stability studies. Mass spectrometry, particularly LC-MS/MS, ensures highly sensitive detection and indispensable in pharmacokinetic and bioanalytical applications. Thus, garenoxacin shows strong compatibility with spectroscopic and chromatographic methods, making it well suited for both qualitative and quantitative analysis. There was no colorimetric methods reported for estimation of garenoxacin was observed and this study is useful for further development of simple analytical methods for the estimation of garenoxacin in bulk and its formulations.

REFERENCES:

- 1. Azhakesannn, Ajitha, and Sujatha Kuppusamy. QbD- Based development and validation of novel stability-indicating method for the assay and dissolution of garenoxacin in garenoxacin tablets. Journal of AOAC international. 2022; 105; 2: 370-378.
- 2. Panchal AB, Patel BR, Patel JG, Patel YK, Maurya UJ. Development and validation of stability indicating
- 3. RP-HPLC method for estimation of Garenoxacin Mesylate in tablet dosage form. International Journal of Pharmacy& Life Sciences. 2017; 8.https://www.ema.eurrupa.eu/en/error/404.
- 4. Edlabadkar AP, Rajput AP. Development of Four Simple UV-Spectrophotometry Methods for Estimation of Garenoxacin Mesylate in Bulk Material and in Pharmaceutical Formulation. Der Pharmacia Letter. 2016;8:213https://www.nhs.uk/c onditions/pages/introduction.aspx.
- 5. Thakur D, Kaur A, Sharma S. Application of QbD based approach in method development of RP-HPLC for simultaneous estimation of antidiabetic drugs in pharmaceutical dosage form. Journal of Pharmaceutical Investigation, 2017;47:229. doi:10.1007/s40005-016-0256-x.

- 6. Woods RK, Dellinger EP. Current guidelines for antibiotic prophylaxis of surgical wounds. Am Fam physician. 1998;57(11):2731-40.
- 7. Supriya GN, Kandisa VR, Ravi Kumar BVV. High performance liquid chromatographic method for determination of garenoxacin mesylate in pharmaceutical dosage form. Journal of Biotechnology Biomater. 2012; 2:194.
- 8. Thakur D, Kaur A, Sharma S. Application of QbD based approach in method development of RP-HPLC for simultaneous estimation of antidiabetic drugs in pharmaceutical dosage form. Journal of Pharmaceutical Investigation, 2017; 47:229. doi:10.1007/s40005-016-0256-x.
- 9. Ishii Y. Antibiotic susceptibility testing and breakpoint. -JPN J Che mother. 2011; 59:454-59. Japanese.
- 10. Garenoxacin NDA accepted for FDA review. It is available at https://www.drugs.com/nda/garenoxaci n_060213.html. [Accessed December 10, 2015].
- 11. Jones RN, Fritsch TR, Sader HS, et al. Garenoxacin (BMS-284756): Invitro antimicrobial spectrum, mechanisms of action, and resistance development. Clinical infectious diseases. 2005; 41 (suppl 2): S87-S98.
- 12. Hooper DC. Mechanisms of action of antimicrobial agents. Principles and practice of infectious diseases, 8th edition. Elsevier, 2015.
- 13. Yamagishi et al. 2015- severe renal failure patients investigated PK of single doses (200-400mg) of garenoxacin.
- 14. Yamagishi, Yuka, Tatsuya Shibata, Satoshi Nakagawa, Junichi Mitsuyama, and Hiroshige Mikamo. Proposed pharmacokinetic and other quipharmacodynamic breakpoint of garenoxacin and other quinolones. Japanese journal of infectious disease. 2017; 70 (6): 616-620.
- 15. Lister PD, Impact of AUC/MIC ratios on the pharmacodynamics of the des-F-6 quinolone garenoxacin is similar to other fluoroquinolones. Journal of

- antimicrobial chemotherapy. 2003; 51: 199-202.
- 16. Ricci, Vito, and Laura Piddock. Accumulation of garenoxacin by bacteroides fragilis compared with that of five fluoroquinolones. Journal of antimicrobial chemotherapy. 2003; 52 (4): 605-609.
- 17. Andrews, J. D. Honeybourne, G. Jevons, M. Boyce, R. WISE, A. Bello and D. Gajjar. Concentrations of garenoxacin in plasma, following a single oral 600mg dose. Journal of antimicrobial chemotherapy. 2003; 51(3): 727-730.
- Krishna, Gopal, Mark H. Gotfried, Kenneth Rolston, and Zaiqi Wang. Penetration of garenoxacin into lung tissues and bones. Current medical research and opinion, 2007; 23 (8): 1841-1847.
- 19. Takagi, Hiroyasu, Kiyoshi Tanaka, Hisatsugu Tsuda. Clinical studies of garenoxacin. International journal of antimicrobial agents. 2008; 32 (6): 468-474.
- 20. Skoog. D. A, Holler. F. G, Nieeman. T.A. principles of instrumental method of analysis. 5th edition, Thomson Brooks Asia. Pvt. Ltd. Singapore. 2004, 4-7.
- 21. Willard. H. H, Meritt, L. Dean, J. A. Settle. Instrumental method of analysis. 7th edition, CBS publishers and distributors, New Delhi, 8-10.
- 22. Chatwal. G. R, Anand. S. K. instrumental method of chemical analysis. 5th edition, Himalaya publications, New Delhi, 2007; 624-2,629-5.
- 23. Sethi. P. High performance liquid chromatography. 1st edition, CBS publishers and distributors, New Delhi, 2001, 12-15.
- 24. National centre for biotechnology information. PubChem compound summary for CID 124093, Garenoxacin. 2023.
- 25. Azhakesannn, Ajitha, and Sujatha Kuppusamy. QbD- Based development and validation of novel stability-indicating method for the assay and dissolution of garenoxacin in

- garenoxacin tablets. Journal of AOAC international. 2022; 105; 2: 370-378.
- 26. D. A. Gajjar. A. Bello, Z. Ge, L. Christopher, and D. M. Grasela, dose safety and pharmacokinetics of garenoxacin, DOI: 10.1128/AAC.47.7.2256-2263.2003 VOL. 47, NO.7.
- 27. Scott Van Wart, Luann Philips, Elizabeth A. Ludwig, Rene Russo, Diptee A. Gajjar, Akintunde Bello, Paul G. Ambrose, Thaddeus H. Grasela, pharmacokinetics and pharmacodynamics of garenoxacin in patients with acquired respiratory infections,
 - DOI:10.1128/AAC.48.12.4766-4777.2004, Vol. 48, no.12.
- 28. Chauhan A, Mittu B, analytical method development and validation: a concise review. J Anal Bioanal Tech. 2015; 6 (1): 1-5.
- 29. Chung C. C, Herman. L, Lee Y.C, Xue-MingZang. Analytical method validation and instrument performance verification. A John Wiley & Sons, Inc., publication, 2004, 11-12.