



STABILITY INDICATING METHOD DEVELOPMENT AND VALIDATION OF DABIGATRAN ETEXILATE IN PHARMACEUTICAL DOSAGE FORMS BY USING RP-HPLC

A. Srivani, Mohammed Amer, A. Shiva Prasad, Chandan Pyne, Mofidul Islam

Department of Pharmaceutical analysis, Pulla Reddy Institute of Pharmacy, Dommadugu Sanga reddy (dist.) Telangana, India.

*Corresponding author E- mail: asrivani1712@gmail.com

ARTICLE INFO

Key words:

Method development ,
Validation , RP-HPLC

Access this article online

Website:

<https://www.jgtps.com/>

Quick Response Code:



ABSTRACT

In the research analysis a rapid, accurate and reliable High Performance Liquid Chromatography (HPLC) method was developed and validated by selecting chromatographic parameters for estimation of Dabigatran etexilate in pharmaceutical dosage forms. The HPLC method was developed using reverse phase PHENOMENEX Luna C₁₈, 5µm, 250 x 4.6mm (size) column with mobile phases containing Methanol (70%) : water (30%). The flow rate was 1.2ml / min with UV-Visible Detector (SPD-20A) detection at λ max 230 nm and the injection volume was set at 20µl with 10min run time. This method has been validated by the use of different validation parameters such as accuracy, precision, linearity, lod and loq. Such findings showed that the system could find practical use in its tablet dosage forms as a quality assurance tool for evaluating the drug in pharmaceutical industries.

INTRODUCTION

Dabigatran etexilate is an oral prodrug that is hydrolyzed to the competitive and reversible direct thrombin inhibitor dabigatran. Dabigatran etexilate may be used to decrease the risk of venous thromboembolic events in patients in whom anticoagulation therapy is indicated. In contrast to warfarin, because its anticoagulant effects are predictable, lab monitoring is not necessary. Dabigatran etexilate was approved by the FDA.

METHOD DEVELOPMENT

HPLC Instrumentation & Conditions

High performance liquid chromatography SHIMADZULC 20AT reciprocating dual pump, UV-Visible Detector (SPD-20A), a reversed-phase PHENOMENEX Luna C₁₈, 5µm, 250 x 4.6mm (size) used for separation. Chromatographic data was acquired using Spin Chrome software & Hamilton injector used.

Standard and sample preparation for UV-spectrophotometer analysis

25 mg of Dabigatran Etexilate Mesylate standard was transferred into 25 ml volumetric flask, dissolve and make up to volume with mobile phase. Further dilution was done by transferring 0.1 ml of the above solution into a 10ml volumetric flask and made up to the volume with mobile phase. The standard and sample stock solutions were prepared separately by dissolving standard and sample in a solvent in mobile phase diluting with the same solvent. (After optimization of all conditions) for UV analysis. It is scanned in the UV spectrum in the range of 200 to 400nm. This has been performed to know the maxima of Dabigatran Etexilate Mesylate, so that the same wave number can be utilized in HPLC-UV detector for estimating the Dabigatran Etexilate Mesylate. While scanning the Dabigatran solution we observed the maxima

at 230nm. The UV spectrum has been recorded on ELICO Double Beam SL210 UV – Vis spectrophotometer.

MOBILE PHASE PREPARATION

Methanol and water were mixed in the ratio of 70:30 to make 1000ml of the mobile phase. It was filtered through 0.45 µm membrane filter and degassed.

SAMPLE & STANDARD PREPARATION FOR THE ANALYSIS

75 mg of Dabigatran Etxilate Mesylate standard was transferred into 75 ml volumetric flask, dissolved and made up to volume with mobile phase. Further dilution was done by transferring 0.2 ml of the above solution into a 10ml volumetric flask and made the required volume with mobile phase.

Table 1: Optimization of chromatographic studies

The chromatographic conditions were optimized by different means. Using different column, different mobile phase, different flow rate, different detection wavelength and different diluents for sample preparation etc.

Column Used	Mobile Phase	Flow Rate	Wave length	Observation	Result
PHENOMENEX Luna C ₁₈ , 5µm, 250 x 4.6mm (size)	ACN : Water = 60 : 40	1 ml/min	230nm	Low response	Method rejected
PHENOMENEX Luna C ₁₈ , 5µm, 250 x 4.6mm (size)	Methanol: Buffer = 70 : 30	1ml/min	230 nm	Very low response	Method rejected
PHENOMENEX Luna C ₁₈ , 5µm, 250 x 4.6mm (size)	ACN: Buffer = 70 : 30	1.0 ml/min	230nm	Tailing peak	Method rejected
PHENOMENEX Luna C ₁₈ , 5µm, 250 x 4.6mm (size)	Methanol: water= 80:20	1.0 ml/min	230nm	Broad Peak	Method rejected
PHENOMENEX Luna C ₁₈ , 5µm, 250 x 4.6mm (size)	Methanol :water = 70:30	1.2ml/min	230nm	Good sharp peak	Method accepted

Table 2: SUMMARY OF OPTIMIZED CONDITIONS

Mobile phase	Methanol :water = 70:30
Wavelength	230 nm
Flow rate	1.2ml/ min.
Run time	10 min.
Column	PHENOMENEX Luna C ₁₈ , 5µm, 250 x 4.6mm (size)

METHOD VALIDATION

1. Accuracy: Recovery study: To determine the accuracy of the proposed method, recovery studies were carried out by adding different amounts (80%, 100%, and 120%) of pure drug of Dabigatran Etxilate Mesylate were taken and added to the pre-analysed formulation of concentration 10µg/ml. From that percentage recovery values were calculated.

2. Precision

Repeatability

The precision of each method was ascertained separately from the peak areas & retention

Times obtained by actual determination of five replicates of a fixed amount of drug. Dabigatran Etxilate Mesylate(API). The percent relative standard deviation was calculated for Dabigatran Etxilate Mesylate are presented.

3. Linearity & Range: The calibration curve showed good linearity in the range of 0 – 25 µg/ml, for Dabigatran Etxilate Mesylate (API) with correlation coefficient (r²) of 0.998. A typical calibration curve has the regression equation of y =70244x + 34176 for Dabigatran Etxilate Mesylate.

4.Limit of Detection: The limit of detection for the drug Dabigatran Etxilate Mesylate can be calculated from the formula

$$LOD = \frac{3.3\sigma}{S}$$

Where, σ = the standard deviation of the response, S = the slope of the calibration curve
The slope S may be estimated from the calibration curve of the analyte. For this method, the value was found to be 0.000451 $\mu\text{g/ml}$ & area 31.76 Dabigatran Etxilate Mesylate.

5. Limit of Quantification: The limit of quantification for the drug Dabigatran Etxilate Mesylate can be calculated from the formula

$$LOQ = \frac{10\sigma}{S}$$

Where, σ = the standard deviation of the response, S = the slope of the calibration curve

The slope S may be estimated from the calibration curve of the analyte. The LOQ for this method was found to be 0.00137 $\mu\text{g/ml}$ & area 96.27 Dabigatran Etxilate Mesylate

6. Robustness:

Chromatographic conditions variation: To demonstrate the robustness of the method, prepared solution as per test method and injected at different variable conditions like using different conditions like flow rate and wavelength. System suitability parameters were compared with that of method precision.

Acceptance criteria: The system suitability should pass as per the test method at variable conditions.

Table 3: Results of Accuracy

Sample ID	Concentration ($\mu\text{g/ml}$)		%Recovery of Pure drug	Statistical Analysis
	Pure drug	Formulation		
S ₁ : 80 %	8	10	96.90	Mean= 96.38% S.D. = 0.171561 % R.S.D.= 0.1015
S ₂ : 80 %	8	10	95.80	
S ₃ : 80 %	8	10	96.30	
S ₄ : 100 %	10	10	96.90	Mean= 96.38% S.D. = 0.045826 % R.S.D.= 0.1015
S ₅ : 100 %	10	10	95.80	
S ₆ : 100 %	10	10	96.30	
S ₇ : 120 %	12	10	96.40	Mean= 96.49% S.D. = 0.105987 % R.S.D. = 0.01
S ₈ : 120 %	12	10	96.50	
S ₉ : 120 %	12	10	96.30	

Table 4: Results of Precision

HPLC Injection Replicates of Dabigatran Etxilate Mesylate	Area	Retention Time
Replicate – 1	2257741	4.63
Replicate – 2	2742277	4.65
Replicate – 3	2741481	4.61
Replicate – 4	2747443	4.62
Replicate – 5	2736501	4.68
Average	2645058	4.63
Standard Deviation	43477.658	0.008944
% RSD	0.01	0.18

Table 5: Results of Linearity and Range

CONC.(µg/ml)	MEAN AUC (n=6)
0	0
5	393853
10	765001
15	1113791
25	1761665

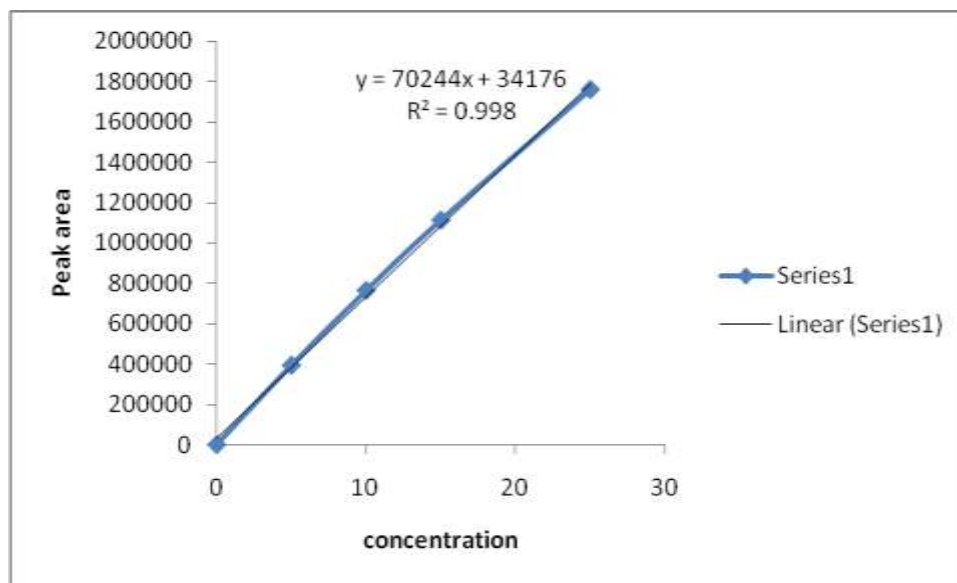


Fig. 1: Linearity Graph

Table 6: Robustness data of the proposed method

VARIATION	RETENTION TIME	AREA	% RSD
FLOW RATE 1ml/min	1	27365014	0.03
	2	27376108	
	3	27365618	
	4	27386816	
	5	27375521	
	6	27385484	
WAVELENGTH 228nm	4.62	2647443	0.02%
	4.64	2646845	
	4.62	2645978	
	4.61	2646654	
	4.61	2646850	
	4.66	2646969	
WAVELENGTH 232nm	4.65	2645841	0.04 %
	4.66	2647540	
	4.65	2646812	
	4.61	2646658	
	4.66	2648534	
	4.68	2645298	

Stability related impurity studies: Following protocol was strictly adhered to for forced degradation of Dabigatran Etxilate Mesylate Active Pharmaceutical Ingredient (API).The API (Dabigatran Etxilate Mesylate) was subjected to stress conditions in various ways to observe the rate and extent of degradation that is likely to occur in the course of storage and/or after administration to body. This is one type of accelerated stability studies that helps us determining the fate of the drug that is likely to happen after long time storage, within a very short time as compare to the real time or long term stability testing. The various degradation pathways studied are acid hydrolysis, basic hydrolysis, thermal degradation and oxidative degradation

Acid hydrolysis: An accurately weighed 10mg. of pure drug was transferred to a clean and dried 10ml volumetric flask. To which 0.1

N Hydrochloric acids was added, volume made up to the mark and kept aside for 24 hrs., from that 0.2 ml was taken in to a 10 ml volumetric flask and volume made the up to the mark with mobile phase, then injected into the HPLC system against a blank of HCl (after all optimized conditions).

Basic hydrolysis: An accurately weighed 10 mg. of pure drug was transferred to a clean and dried 10 ml volumetric flask. To which 0.1 N Sodium hydroxide was added and volume made up to the mark and kept aside for 24 hrs., from that 0.2 ml was taken in to a 10 ml volumetric flask and volume made up to the mark with mobile phase, then injected into the HPLC system against a blank of 0.1NaOH (after all optimized conditions).

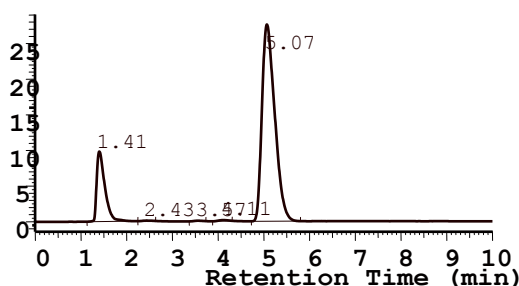


Table 7: Results of forced degradation studies of Dabigatran Etxilate Mesylate API

Stress condition	Time	Assay of active substance	Assay of degraded products	Mass Balance (%)
Acid Hydrolysis (0.1 N HCl)	24Hrs.	98.36	-----	98.36
Basic Hydrolysis (0.1N NaOH)	24Hrs.	98.32	-----	98.32
Oxidation (3% H ₂ O ₂)	24 Hrs	97.31	-----	97.31

RESULTS AND DISCUSSION

To develop a precise, linear, specific and suitable stability indicating RP-HPLC method for analysis of Dabigatran Etxilate Mesylate, different chromatographic conditions were applied and the results observed are presented in previous chapters.

Isocratic elution is simple, requires only one pump and flat baseline separation for easy and reproducible results. So, it was preferred for the current study over gradient elution. Mobile phase and diluent for preparation of various samples were finalized after studying the solubility of API in different solvents of our disposal (acetonitrile, methanol, water and phosphate buffer).

The drug was found to be highly soluble in methanol. Drug was insoluble in water and acetonitrile. Using these solvents with appropriate composition newer method can be developed and validated. Detection wavelength was selected after scanning the standard solution of drug over 200 to 400nm. From the U.V spectrum of Dabigatran Etxilate Mesylate it is has observed λ max 230 nm. Further, a flow rate of 1.2 ml/min and an injection volume of 20 μ l were found to be the best analysis. The result showed the developed method is yet another suitable method for assay and stability studies which can help in the analysis of Dabigatran Etxilate Mesylate in different formulations. The method can also be used for get stability related impurities in Dabigatran Etxilate Mesylate. There are four impurity peaks has been seen in acidic condition, two in basic.

CONCLUSION

A sensitive & selective RP-HPLC method has been developed & validated for the analysis of Dabigatran Etxilate Mesylate API. Further the proposed RP-HPLC method has excellent sensitivity, precision and reproducibility. The result shows the developed method is yet another suitable method for assay, impurity studies which can help in the analysis of Dabigatran Etxilate Mesylate in different formulations

REFERENCES

1. Parimoo P. Pharmaceutical analysis. CBS Publishers and Distributors. New Delhi, 145.
2. Mendham J, Denny RC and Thomas M. 2004. Vogel's textbook of quantitative analysis. 6th ed., Pearson Education Limited.
3. Sharma BK. Instrumental methods analysis. 17th ed. Goel publishing house, Meerut, 44.
4. Willard HH, Merritt LL (Jr.), Dean JA, SettleFFA (Jr.). 1986. Instrumental methods of chemical analysis. 6thed. New Delhi: CBS Publishers.
5. Connors KA. 1994. A Textbook of pharmaceutical analysis. 3rd ed. Delhi. Wiley inter sciences Inc.
6. Alsante K M, Boutres P, Couturier M A. 2004. Pharmaceutical Impurity Identification: A Case Study Using a Multidisciplinary Approach. Journal of Pharmaceutical Sciences. 93 (9). 2296.
7. International Conference on Harmonization. 2000. Draft Revised Guidance on Impurities in New Drug Substances. Federal Register Q3A(R). 65 (140). 45085.
8. International Conference on Harmonization. 2000. Draft Revised Guidance on Impurities in New Drug Products. Federal Register Q3B(R). 65 (139). 44791.
9. International Conference on Harmonization Impurities. 1997. Q3C-Guidelines for Residual Solvents. Q3C, Federal Register. 62 (247). 67377.
10. International Conference on Harmonization Specifications.1999. Q6A: Test Procedures and Acceptance Criterial for New Drug Substances and New Drug Products. Chemical substances. 65 (146). 67488.
11. Alsante K M, Hatajik T D, Lohr L L and Sharp T R. 2001. Isolation and Identification of Process Related Impurities and Degradation Products from Pharmaceutical Drug Candidates. Part 1. American Pharmaceutical Review. 4(1). 70.
12. Lohr L L, Sharp T R, Alsante K M and Hatajik T D. 2001. Isolation and Identification of Process Related Impurities and Degradation Products from Pharmaceutical Drug Candidates. Part II: The Roles of NMR and Mass Spectrometry. American Pharmaceutical Review. Fall issue.
13. Winger B E, Kemp C A. 2001. Characterization of Pharmaceutical Compounds and Related Substances by

- using FTICR-MS and Tandem Mass Spectrometry. American Pharmaceutical Review. Summer issue.
14. ICH Topic Q3 A. 1995. Impurities Testing Guideline: Impurities in New Drug Substances, The European Agency for the Evaluation of Medicinal Products Human Medicines Evaluation Unit.
 15. Farmer S, Anderson P, Burns P and Velagaleti R. 2002. Forced Degradation of Ibuprofen in Bulk Drugs and Tablets. *Pharmaceutical Technology*. 28. 42.
 16. Bhat P and Velingkar V S. 2004. Synthesis and Characterization of Degradation Products in Diclofenac-sodium and Clotrimazole. *Indian Drugs*. 40 (7). 396.
 17. Volk K J, Hill S E, Kerns E H, Lee M S. 1997. Profiling degradants of paclitaxel using liquid chromatography-mass spectrometry and liquid chromatography-tandem mass spectrometry sub structural techniques. *Journal of Chromatography*. 696(1).99.
 18. Riley T N. 1998. steric aspects of drug action. *Pharmacist*. 23 (3). 40.
 19. Jacobs P, Dewe W, Flament A, Gibella M, Ceccato A. 2005. a new validation approach applied to the GC determination of impurities in organic solvents. *Journal of Pharmaceutical and Biomedical analysis*. 40. 294.
 20. Jack Yuk K Cheng, Man Fai Chan, Tai Wai Chan, Mei Yuen Hung. 2006. Impurity profiling of ecstasy tablets seized in Hong Kong by GC-MS. Article in Press *Forensic Science International*. 144. 21.
 21. Gimeno P, Besacier F, Bottex M, Dujourdy L, Chaudron-Thozet H. 2005. A study of impurities in intermediates and 3, 4 methylenedioxymethamphetamine (MDMA) samples produced via reductive amination routes. *Forensic Science International*. 155. 141.
 22. Buhler V. 1998. *Vademecum for Vitamin Formulation*. Verl-Ges. 142. 36.
 23. Hoq M M, Morsheda S B and Gomes D J. 1991. Development of appropriate preservative system for liquid antacid: bacterial contaminants in antacid samples. *Journal of Microbiology*. 8(1), 5.
 24. Roy J, Islam M, Khan A H, Das S C, Aktheruzzaman M, Deb A K, Alam A H. 2001. Diclofenac sodium injection sterilized by autoclave and the occurrence of cyclic reaction producing a small amount of impurity. *Journal of Pharmaceutical Sciences*. 90. 541.
 25. Roy J, Mahmud M, Sobhan A, Aktheruzzaman M, Al-Faoouque M and Ali E. 1994. Marketed vitamin B-complex injectables: stability and mutual interaction. *Drug Development and Industrial Pharmacy*. 20 (13). 2157.